

Cytokine Secretion Depends on Gal α (1,3)Gal Expression in a Pig-to-Human Whole Blood Model¹

Marit Sæthre,* Mårten K. J. Schneider,[†] John D. Lambris,[‡] Paola Magotti,[‡] Guttorm Haraldsen,[§] Jörg D. Seebach,[¶] and Tom E. Mollnes^{2*}

Transplants from α 1,3-galactosyltransferase (Gal) gene-knockout pigs to nonhuman primates are largely protected from hyperacute but not acute humoral xenograft rejection. The present study investigates the role of Gal in cytokine responses using a novel pig-to-human whole blood in vitro model, developed for species-specific analysis of porcine and human cytokines. Porcine ($n = 7$) and human ($n = 27$) cytokines were measured using ELISA or multiplex technology, respectively. Porcine aortic endothelial cells from control (Gal^{+/+}) and Gal-deficient (Gal^{-/-}) pigs were incubated with human lepirudin anticoagulated whole blood from healthy donors. E-selectin expression was measured by flow cytometry. The C3 inhibitor compstatin and a C5aR antagonist were used to study the role of complement. Cytokine species specificity was documented, enabling detection of 2 of 7 porcine cytokines and 13 of 27 human cytokines in one single sample. Gal^{+/+} porcine aortic endothelial cells incubated with human whole blood showed a marked complement C5b-9 dependent up-regulation of E-selectin and secretion of porcine IL-6 and IL-8. In contrast, Gal^{-/-} cells responded with E-selectin and cytokine expression which was so weak that the role of complement could not be determined. Human IL-6, IL-8, IFN- γ , MIP-1 α , MIP-1 β , eotaxin, and RANTES were detected in the Gal^{+/+} system, but virtually no responses were seen in the Gal^{-/-} system ($p = 0.03$). The increase in human cytokine release was largely complement dependent and, in contrast to the porcine response, mediated through C5a. Species-specific analysis of cytokine release revealed a marked, complement-dependent response when Gal^{+/+} pig cells were incubated with human whole blood, compared with Gal^{-/-} cells which induced virtually no cytokine release. *The Journal of Immunology*, 2008, 180: 6346–6353.

Xenotransplantation holds promise for unrestricted supply of organs, but several immunological hurdles must be overcome before xenotransplantation can become a clinical reality. Pigs are currently considered the donor of choice for human recipients. Nevertheless, discordant xenotransplantation, e.g., pig to primate, induces strong immune responses that lead to rapid loss of the graft. The primary hyperacute rejection (HAR)³ in human and nonhuman primates is caused by naturally occurring Abs that bind to the carbohydrate epitope Gal α 1,3Gal (Gal) on pig

vascular endothelium and subsequently activate the complement system (1–4). Accordingly, HAR can be prevented by using pig organs transgenic for complement regulators (5, 6) or deficient for Gal by targeted deletion of the α 1,3-galactosyltransferase (*GalT*) gene (7, 8). If HAR is inhibited, organs may subsequently be rejected in a process called acute humoral xenograft rejection (AHXR), also termed acute vascular rejection (9, 10). The pathogenesis of AHXR is not fully understood and organs from Gal-deficient pigs are still rejected by AHXR when transplanted into nonhuman primates (11, 12), and Abs against non-Gal epitopes may partly mediate this rejection (13–15).

Currently, a lot of attention is paid to the mechanism of rejection in the absence of Gal. In the present study, we have extended our in vitro model to include incubation of porcine aortic endothelial cells (PAEC) with human whole blood instead of serum (16). Thus, the effects of such coculture on both the human blood and the PAECs were explored. To this end, we have focused on establishing the cytokine profile of both the porcine endothelium and human blood by using species-selective assays. The role of cytokines in xenotransplantation is interesting given their well-established role in the pathogenesis of allograft rejection (17, 18), and numerous reports describing elevated levels in xenotransplantation (19, 20). However, their dependence on Gal-mediated immune activation upon exposure of xenograft endothelium has not been explored. Human polymorphonuclear cells (PMN) represent a major source of proinflammatory factors, including TNF- α , IL-1, IL-8, MIP-1 α , and MIP-1 β , and recruit additional PMN to the sites of inflammation (21, 22). PAECs also induce IL-1 β , PGE₂, and TNF- α production in human monocytes (23).

There are different roles for Gal in direct human leukocyte interactions with porcine endothelial cells (EC). Gal on porcine EC constitutes potential binding sites for human PMN, NK cells, and monocytes. Human NK cells may induce porcine EC activation by

*Institute of Immunology, Rikshospitalet University Hospital and Faculty of Medicine, University of Oslo, Oslo, Norway; [†]Laboratory of Transplantation Immunology, Department of Internal Medicine, University Hospital, Zurich, Switzerland; [‡]Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia, PA 19104; [§]Laboratory for Immunohistochemistry and Immunopathology, Division of Pathology, Rikshospitalet University Hospital and Faculty of Medicine, University of Oslo, Oslo, Norway; and [¶]Service of Immunology and Allergology, Department of Internal Medicine, Geneva University Hospital and Faculty of Medicine, Geneva, Switzerland

Received for publication January 3, 2008. Accepted for publication March 3, 2008.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

¹ This work was supported by The Norwegian Research Council, The Research Council of Rikshospitalet, The Norwegian Council on Cardiovascular Disease, The Family Blix Foundation, The Odd Fellow Foundation, Swiss National Science Foundation Research Grant 3200B0-109921, and National Institutes of Health Grants GM-62134 and AI-068730.

² Address correspondence and reprint requests to Dr. Tom Eirik Mollnes, Institute of Immunology, Rikshospitalet University Hospital, N-0027 Oslo, Norway. E-mail address: t.e.mollnes@medisin.uio.no

³ Abbreviations used in this paper: HAR, hyperacute rejection; Gal, Gal α 1-3Gal; AHXR, acute humoral xenograft rejection; PAEC, porcine aortic endothelial cell; KO, knockout; PMN, polymorphonuclear cell; EC, endothelial cell; MFI, median fluorescence intensity; FGF, fibroblast growth factor; PDGF, platelet-derived growth factor; IP-10, IFN- γ -inducible protein 10; VEGF, vascular endothelial growth factor.

Copyright © 2008 by The American Association of Immunologists, Inc. 0022-1767/08/\$2.00

direct interaction in the absence of Gal (24, 25) and no significant reduction of direct NK cytotoxicity or NK cell and PMN adhesion on porcine ECs was found in the absence of, or upon blocking, Gal. In contrast, monocyte adhesion to porcine ECs depends on Gal-mediated activation (26).

In a recent study (16), we observed a marked reduction in human serum-induced E-selectin (CD62E) expression in PAECs lacking the Gal (Gal^{-/-}) when compared with normal (Gal^{+/+}) cells. In the present study, the mutual inflammatory response of the PAECs and the human blood cells was studied by establishing a species-specific detection system to measure porcine and human mediators in the same sample. Furthermore, specific complement inhibitors were used to evaluate the role of complement in this response.

Materials and Methods

Cell culture

Primary PAECs from GalT-knockout (KO; Gal^{-/-} PAECs) or control littermate (Gal^{+/+} PAECs) pigs were provided by R. J. Hawley (formerly Immerge Biotherapeutics), and used up to passage number 20. The immortalized PAEC line PEDSV.15 (Gal^{+/+}) was generated as previously described in detail (27). The Gal^{-/-} cell line PED2*3.51 was generated from PEDSV.15 by homologous recombination, panning, and limiting dilution cloning (28). All cells were cultured in DMEM (Invitrogen Life Technologies) supplemented with 10% FCS (BioWhittaker), 1 mM sodium pyruvate, 2 mM L-glutamine, 0.1 mM nonessential amino acids, 20 mM HEPES, 50 µg/ml gentamicin (all obtained from Invitrogen Life Technologies), and 0.25 µg/ml amphotericin B (BioWhittaker).

Human whole blood

Blood from five healthy volunteers was collected into sterile polypropylene tubes (4.5-ml Nunc cryotubes; Nalgene International) containing 50 µg/ml lepirudin (refludan; Hoechst) as anticoagulant. Samples were obtained immediately before each experiment and kept on ice. Informed written consent was obtained from each donor. The study was approved by the local ethical committee.

In vitro pig-to-human transplantation model

Gal^{+/+} and Gal^{-/-} PAECs (both primary and immortalized) were grown to confluence in 96-well cell culture plates (Corning). The cells were then exposed to 100 µl of human whole blood, diluted to 50% in culture medium, for 24 h on a shaker in a CO₂ incubator. After incubation, cell supernatants were collected, centrifuged, and immediately frozen at -70°C. PAECs were then washed, trypsinized, washed with medium containing 10% FCS, and centrifuged at 300 × g for 5 min. PE-conjugated anti-CD62E (E-selectin) clone 1.2B6 or PE-conjugated isotype control IgG1 (both obtained from Southern Biotechnology Associates) were added directly to the pellet and cells were incubated for 30 min at 4°C, and subsequently analyzed by flow cytometry (FACScan; BD Biosciences). Cells incubated with TNF-α (Sigma-Aldrich) were used as positive control. Results are given as median fluorescence intensity (MFI).

Porcine cytokine measurement

Pilot samples from the PAEC-human whole blood model were examined in a panel of seven porcine cytokine ELISAs (TNF-α, IL-1β, IL-6, IL-8, IL-10, IL-12(p70), IFN-γ), performed according to the instructions from the manufacturer (R&D Systems). The porcine ELISAs are quantitative sandwich immunoassays, precoated with the actual anti-porcine cytokine Ab (usually monoclonal). Briefly, standards and samples were added to the wells. After washing, an enzyme-linked Ab (usually polyclonal) reacting with the porcine cytokine was added. Following a final wash, the plates were subsequently developed with substrate solution for 30 min at room temperature, stop solution was added, and OD was measured at 405 nm using a microplate reader (Asys Hightech). Data were analyzed with MicroWin software (Mikrotek Laborsysteme). A control sample with known concentrations of the porcine cytokine was included on every plate.

Human cytokine measurement

Multiplex technology was used to measure the concentration of a broad panel of human cytokines, chemokines, and growth factors (Human Cytokine 27-plex-kit; Bio-Rad). This is a bead-based sandwich immunoassay, which simultaneously measure the following 27 human biomarkers: IL-1β,

Table I. *Species specificity in porcine inflammatory mediator assays^a*

Mediator	Human Plasma	Porcine Plasma	Detected in the Present Model
IL-1β	22	9,000	
IL-6	0	150	X
IL-8	0	60,000	X
IL-10	0	700	
IL-12(p70)	0	500	
IFN-γ	0	40	
TNF-α	0	10,000	

^a Plasma obtained from human and porcine blood incubated with Gram-negative bacteria examined in ELISAs for detection of porcine inflammatory mediators (values are given in picograms per milliliter). Cytokines marked X (right column) were detected in the present pig-to-human in vitro whole blood model using both primary and immortalized PAECs.

IL-1ra, IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12(p70), IL-13, IL-15, IL-17, TNF-α, IFN-γ, IFN-γ-inducible protein 10 (IP-10), fibroblast growth factor (FGF)-basic, MIP-1α, MIP-1β, MCP-1, G-CSF, GM-CSF, platelet-derived growth factor (PDGF)-BB, eotaxin, RANTES, and vascular endothelial growth factor (VEGF). The analyses were performed according to instructions from the manufacturer. Briefly, beads of defined spectral properties are conjugated to bind specific capture Abs. The biomarker in each sample will bind to the capture Abs on the beads. Then biomarker-specific biotinylated detector Abs added will bind to the appropriate immobilized analytes. Finally, streptavidin conjugated to the fluorescent protein R-PE binds to the biotinylated detector Abs associated with the immune complexes of the beads, forming a solid-phase sandwich. The beads were analyzed with a Luminex 100, Bio-Plex array reader (Bio-Rad). Standard curves were generated for each biomarker using the mixed analyze standard provided with the kit.

Species specificity of the porcine and human cytokine detection assays

The experimental model of PAECs incubated with human whole blood requires that the cytokines in the plasma supernatant are detected in species-specific assays for correct interpretation of the results. Human whole blood was incubated with *Neisseria meningitidis* bacteria (strain H44/76, isolated from a patient with invasive meningococcal disease (29)), and porcine whole blood was incubated with *Escherichia coli* (American Type Culture Collection) (both 10⁸ bacteria/ml blood) to produce plasma samples with large amounts of human and porcine cytokines, respectively (T. E. Mollnes et al., unpublished data). These two samples were tested in both species assays described above. Based on the high specificity of these assays (Table I), the current study was undertaken.

Complement inhibitors

Compstatin is a 13-aa cyclic peptide that binds and inhibits cleavage of complement component C3 (30, 31). The sequence is Ac-I[CV(1MeW)QDWGAHRC]T and the molecular mass is 1628 Da. A linear-inactive analog to compstatin, differing in only one amino acid, was used as a control. The sequence is Ac-IAVVQDWGHHHRAT and the molecular mass is 1532 Da. To block complement C5aR on leukocytes, a small cyclic hexapeptide that acts as a selective C5aR antagonist (C5aRa) was used (32, 33). The sequence is AcF-[OPdChaWR] and the molecular mass is 896 Da. The inhibitors and the control peptide were produced in the laboratory of J. D. Lambris.

Statistical analysis

The Wilcoxon test for paired samples was used. Values of *p* < 0.05 were considered statistically significant.

Results

E-selectin expression is strongly up-regulated on Gal^{+/+}, but not Gal^{-/-}, PAECs incubated with human whole blood

TNF-α, used as positive control, caused a marked up-regulation of E-selectin on both Gal^{+/+} (220 MFI) and Gal^{-/-} (205 MFI) immortalized PAECs compared with resting cells (29 and 19 MFI, respectively) (Fig. 1). Human serum induced a substantial up-regulation of E-selectin on Gal^{+/+} PAECs, whereas virtually no effect

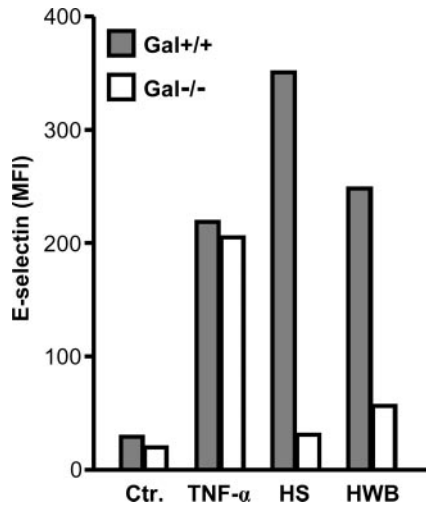


FIGURE 1. E-selectin expression on immortalized Gal^{+/+} and Gal^{-/-} PAECs. PAECs were incubated 4 h with TNF- α , 50% human serum (HS), and 50% human whole blood (HWB). E-selectin cell surface expression was measured by flow cytometry. Data represent MFI in one representative experiment.

was seen on Gal^{-/-} cells (351 and 31 MFI, respectively). The effect of human whole blood on porcine E-selectin expression was comparable to the response seen with serum alone (248 MFI on Gal^{+/+} and 56 MFI on Gal^{-/-} cells) (Fig. 1), indicating that plasma factors rather than blood cells were the main inducers of this effect. No staining of the cells was observed using an isotype-matched control Ab.

Human blood-induced E-selectin up-regulation is reversed by complement inhibition

The increase in E-selectin on immortalized Gal^{+/+} cells was dose-dependently and completely reduced by the C3 inhibitor compstatin, whereas the C5aR antagonist and the control peptide had no effect (Fig. 2, closed circles), consistent with a C5b-9-mediated effect in the whole blood model as previously shown for serum (16). By contrast, the minor up-regulation of E-selectin on Gal^{-/-} cells incubated with human whole blood was not affected by any of the complement inhibitors (Fig. 2, open circles).

Species-specific detection of porcine and human inflammatory mediators

Porcine and human plasma was obtained from whole blood incubated with Gram-negative bacteria. These plasma samples were

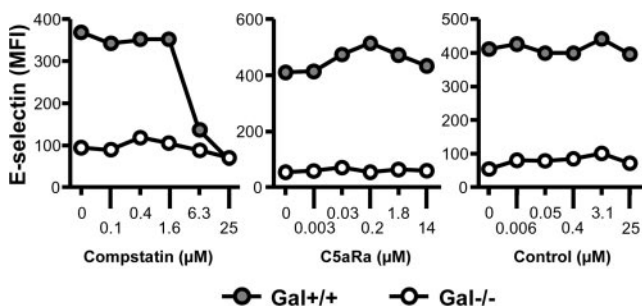


FIGURE 2. Effect of complement blocking on E-selectin expression on immortalized Gal^{+/+} and Gal^{-/-} PAECs. Gal^{+/+} (●) and Gal^{-/-} (○) PAECs were incubated with human whole blood and increasing concentrations of the complement-inhibitors compstatin (left panel) and a C5aR antagonist (middle panel) as well as a control peptide (right panel). E-selectin expression on incubated PAEC was determined by flow cytometry. Representative data from one blood donor are presented.

Table II. Species specificity of human inflammatory mediator assays^a

Mediator	Human Plasma	Porcine Plasma	Detected in the Present Model
IL-1 β	3,480	0	
IL-1ra	3,800	0	X
IL-6	17,000	0	X
IL-8	14,000	0	X
IL-9	80	0	X
IL-10	77	0	
Eotaxin	113	0	X
FGF-basic	131	0	X ^b
G-CSF	180	0	
IFN- γ	460	0	X ^c
IP-10	1,400	0	X
MCP-1	191	196	X
MIP-1 α	6,800	0	X
MIP-1 β	15,000	0	X
PDGF-bb	12,200	2,700	X
RANTES	29,000	223	X
TNF- α	40,000	0	X ^c
VEGF	112	0	X ^c

^a Plasma obtained from human and porcine blood incubated with Gram-negative bacteria (same samples as presented in Table I) examined in a multiplex assay for detection of human inflammatory mediators (values are given in picograms per milliliter). Cytokines marked X (right column) were detected in the present pig-to-human in vitro model using both primary and immortalized PAECs.

^b Detected only using primary cells.

^c Detected only using immortalized cells.

already known to contain high amounts of a number of cytokines (see *Materials and Methods*). To document species specificity of the assays used in this study, the samples were first tested in seven porcine ELISAs (Table I, left and middle columns). Reactivity was found for all of these mediators in porcine plasma. Six of the seven mediators were completely negative when human plasma, known to contain high amounts of the corresponding human variants, were tested, whereas IL-1 β showed a weak reactivity (22 pg/ml in human plasma compared with 9000 pg/ml in porcine plasma). When the same samples were tested in a multiplex assay for detection of human inflammatory markers, reactivity was seen for 18 of 27 markers in human plasma (Table II, left and middle columns). Fifteen of the 18 were completely negative when porcine plasma was tested in the assay, whereas MCP-1, PDGF-bb, and RANTES showed various degree of cross-reactivity. Because species specificity requires that the mediator in question is documented to be present in both the porcine and the human plasma sample, whereas reactivity is found only in the corresponding assay, species specificity was concluded for IL-6, IL-8, IL-10, IFN- γ , and TNF- α . In addition, IL-1 β was highly specific in the human assay and virtually specific in the porcine assay.

Cytokines and chemokines released upon coculture of human whole blood with PAEC

The incubation with human whole blood induced the secretion of porcine IL-6 and IL-8 in both immortalized and primary Gal^{+/+} PAECs (Table I, right column). In the human cells, IL-1ra, IL-6, IL-8, IL-9, IP-10, MIP-1 α , MIP-1 β , MCP-1, PDGF-BB, eotaxin, and RANTES secretion was induced by both immortalized and primary Gal^{+/+} PAEC. In contrast, human TNF- α , IFN- γ , and VEGF were only detected using immortalized cells, whereas FGF-basic was only induced using primary cells (Table II, right column).

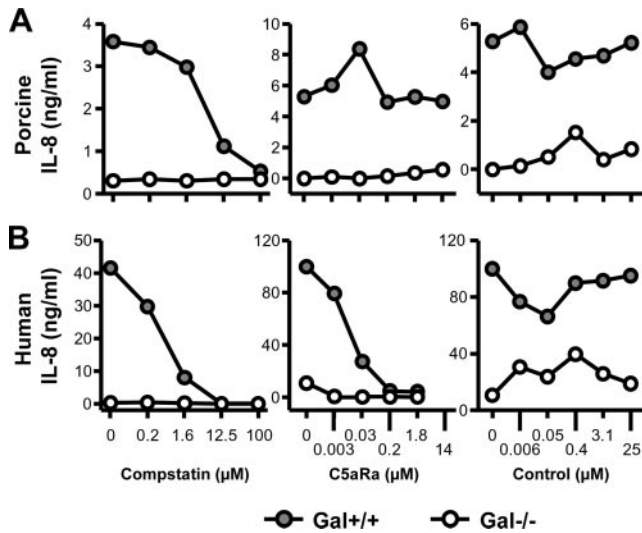


FIGURE 3. Effect of complement blocking on porcine and human cytokine production. Immortalized Gal^{+/+} (●) and Gal^{-/-} (○) PAECs were incubated with human whole blood to measure porcine (A) or human (B) IL-8 production. Complement inhibitors compstatin (left panel) and a C5aR antagonist (middle panel) as well as a control peptide (right panel) were added in increasing concentrations (x-axis). Representative data from one blood donor are presented. Cytokine data are presented as concentration (nanograms per milliliter).

Dose-dependent effect of complement inhibition on porcine and human cytokine release

The differential effect of complement on porcine and human cytokine release as described above was further investigated in dose-response experiments using human whole blood and immortalized PAEC with IL-8 as a readout. In contrast to Gal^{+/+} cells, Gal^{-/-} cells induced neither porcine nor human IL-8 (Fig. 3). Human IL-8 secretion decreased dose-dependently to nadir by both the C3 inhibitor compstatin and a C5aR antagonist (Fig. 3, lower panel). In contrast, porcine IL-8 production by Gal^{+/+} PAEC was reduced to nadir with increasing concentrations of compstatin, whereas no effect was seen using the C5aR antagonist or the control peptide (Fig. 3, upper panel). Accordingly, human whole blood incubation

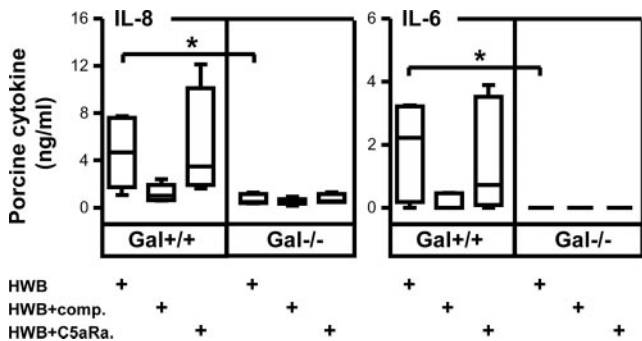


FIGURE 4. Detection of porcine cytokines in cell supernatants. Porcine IL-8 (left panel) and IL-6 (right panel) concentrations were analyzed in supernatants from immortalized Gal^{+/+} and Gal^{-/-} PAECs incubated with human whole blood (HWB) alone, HWB with complement inhibitor C5aR (C5aRa, 14 μM), or HWB with complement inhibitor compstatin (comp., 100 μM). Both IL-8 and IL-6 concentration was significantly higher from Gal^{+/+} compared with Gal^{-/-} porcine PAECs (*, p = 0.03). Data are presented as box plots showing median, quartiles, and minimum and maximum values from five experiments, each using a different donor.

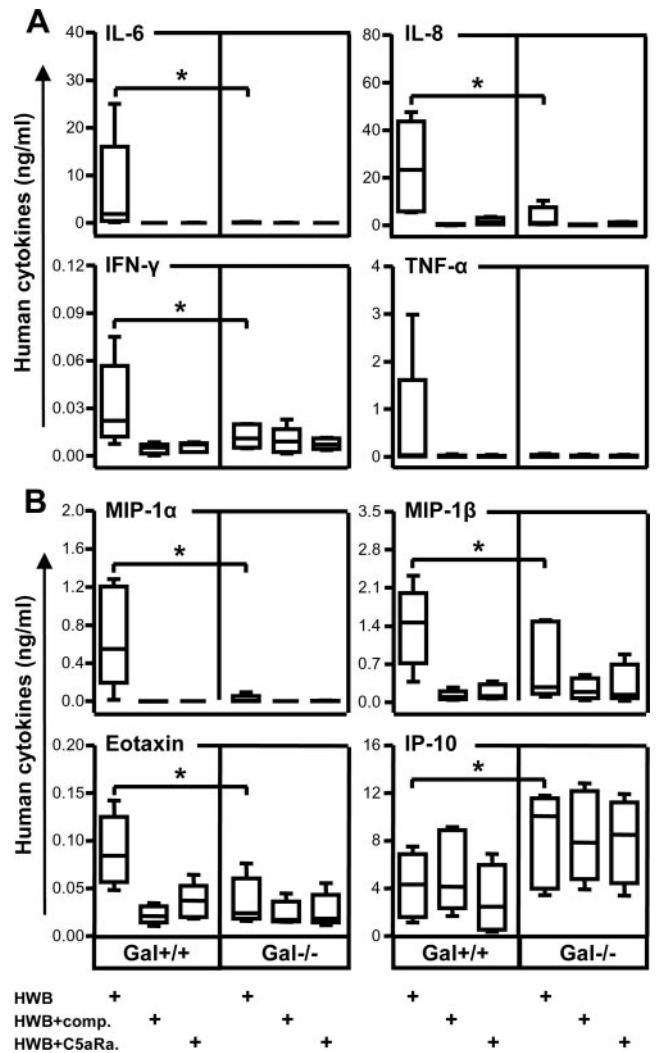


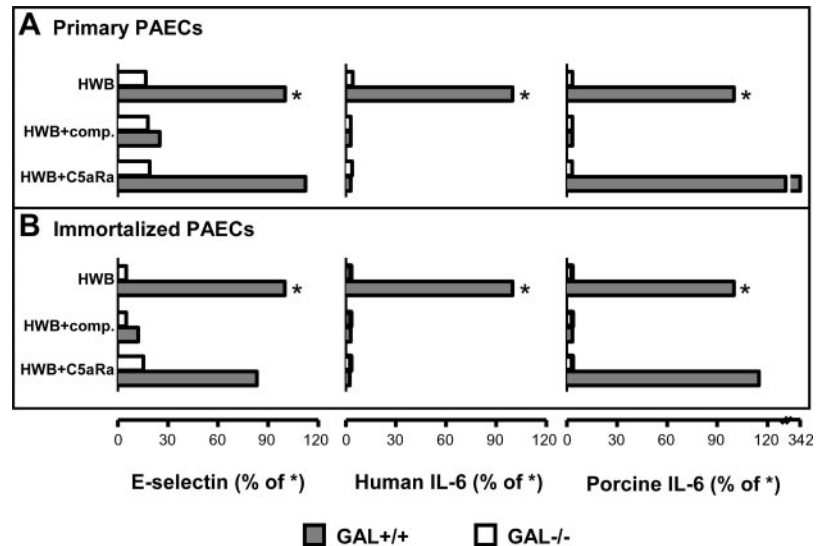
FIGURE 5. Detection of human cytokines in cell supernatants. Cytokine production was detected in cell supernatants when immortalized Gal^{+/+} and Gal^{-/-} PAECs were incubated with human whole blood (HWB) alone or with complement inhibitors C5aR antagonist (C5aRa, 14 μM) or compstatin (comp., 100 μM). A, Human cytokines (IL-6, IL-8, IFN-γ, and TNF-α) for which cross-reactivity to porcine cytokines was excluded in the human multiplex system (Table II). B, Human cytokines (MIP-1α, MIP-1β, eotaxin, and IP-10) for which cross-reactivity with porcine cytokines could not be determined. Significant differences were found between human cytokines detected in the Gal^{+/+} compared with Gal^{-/-} PAECs for all cytokines (*, p = 0.03). Data are presented as box plots showing median, quartiles, and minimum and maximum values from five experiments, each using a different donor.

induced a significant IL-6 production in Gal^{+/+}, but not Gal^{-/-}, PAECs, which was virtually abolished by compstatin but not by the C5aR antagonist (Fig. 4). Thus, complement is essential for both the porcine and the human response, but the effector mechanisms are strikingly different.

Differential induction of human inflammatory mediators by immortalized Gal^{+/+} and Gal^{-/-} PAECs

Significantly higher concentrations of human IL-6, IL-8, and IFN-γ were detected after whole blood incubation with Gal^{+/+} PAECs as compared with Gal^{-/-} cells, whereas the difference for TNF-α was not statistically significant due to large interindividual variation (Fig. 5A). For the chemokines where cross-reactivity could not be determined, significantly higher concentrations of the

FIGURE 6. Comparison of immortalized and primary $\text{Gal}^{+/+}$ and $\text{Gal}^{-/-}$ PAECs. Primary (A) and immortalized PAECs (B) were compared with respect to E-selectin expression and human and porcine cytokines in cell supernatants (here shown as IL-6) after incubation with human whole blood (HWB), HWB with compstatin (Comp.), or HWB with a C5aR antagonist (C5aR). E-selectin expression data represents percent median fluorescent intensity of $\text{Gal}^{+/+}$ PAECs incubated with HWB (marked with an asterisk (*)). Cytokine data are presented as concentration (nanograms per milliliter).



chemokines MIP-1 α , MIP-1 β , and eotaxin were measured in cell supernatants from cultures with $\text{Gal}^{+/+}$ compared with $\text{Gal}^{-/-}$ PAECs (Fig. 5B). In contrast significantly higher concentrations of IP-10 were recorded in cell supernatants from $\text{Gal}^{-/-}$ PAECs as compared with $\text{Gal}^{+/+}$ cells (Fig. 5B, lower right panel). For IL-9, VEGF, and IL-1ra, no difference was seen between $\text{Gal}^{+/+}$ and $\text{Gal}^{-/-}$ PAEC cultures (data not shown). Of the mediators with a certain degree of documented species cross-reactivity, RANTES was significantly higher in the supernatant from $\text{Gal}^{+/+}$ than from $\text{Gal}^{-/-}$ cells, whereas no difference were seen for PDGF-bb and MCP-1 (data not shown). In line with its effect on human IL-8, compstatin virtually abolished the production of IL-6, IL-8, IFN- γ , TNF- α , MIP-1 α , MIP-1 β , and eotaxin (Fig. 5). Moreover, in contrast to the production of porcine cytokines, the C5aR antagonist efficiently inhibited all human cytokines except IP-10 (Fig. 5), again indicating that the cytokine production by human whole blood induced by $\text{Gal}^{+/+}$ cells was C5a mediated. In contrast, the IP-10 production was complement independent.

Comparable results using immortalized and primary PAECs in cocultures with whole human blood

The validity of using the immortalized $\text{Gal}^{+/+}$ PAECs line PESDV.15 and the corresponding $\text{Gal}^{-/-}$ variant has been questioned, e.g., with respect to expression of VCAM-1 (24). We therefore performed a series of coculture experiments using primary PAECs obtained from GalT-KO pigs or their native control animals, and the immortalized lines described above. Primary $\text{Gal}^{+/+}$ and $\text{Gal}^{-/-}$ PAECs incubated with human whole blood showed E-selectin up-regulation and induced IL-6-responses virtually identical with their corresponding immortalized cell lines (Fig. 6). In addition, the effect of compstatin and the C5aR antagonist was equal in the primary and immortalized cell cultures.

Discussion

In the present study, we explored the importance of Gal and non-Gal epitopes in the inflammatory reaction induced when PAECs are exposed to human whole blood, an issue which has become particularly important after the advent of cloned GalT-KO pigs (34, 35). Organs from GalT-KO animals are still rejected by AHXR and show features of thrombotic microangiopathy. Possible explanations for these later rejection processes include molecular incompatibility between pig and primates (36, 37), T cell-independent mechanisms due to natural non-Gal Abs, or T cell-dependent

mechanisms due to elicited non-Gal Abs (38). Growing evidence supports a close relation between inflammation and hemostasis, e.g., as illustrated by the cross-talk between complement and coagulation (39). Thus, control of the inflammatory reaction, as reflected by cytokine release, is required to increase the compatibility between pig and primate organs.

Differences in cytokine production during rejection of cells and organs from GalT-KO compared with wild-type pigs are largely unknown. To investigate this, we used an established human whole blood model (40) to activate $\text{Gal}^{+/+}$ and $\text{Gal}^{-/-}$ PAECs and documented large differences between $\text{Gal}^{+/+}$ and $\text{Gal}^{-/-}$ PAECs with respect to production of both endogenous porcine cytokines and induction of cytokine production from human blood cells. This model is based on anticoagulation with lepirudin, a recombinant hirudin analog which is a highly specific thrombin inhibitor that does not influence complement activation, in contrast to heparin and other available anticoagulants (40). Establishing a whole blood model without using an anticoagulant is complicated, and attempts show that blood coagulates within 2 h (41). Because a time period of several hours is required to study cytokine production, anticoagulation is mandatory and despite inhibition of the last step of coagulation (thrombin), to our knowledge, the protocol used herein represents the most physiological relevant model to study whole blood in vitro. There are no data published on differences between the different endothelial cells used in this study with respect to expression of thrombin receptors or their response to lepirudin. This is a limitation of the current study. Comparing the consistent data obtained from the different cell lines we used, however, there is no indication that lepirudin influenced the results.

The model was extended to include primary PAECs from $\text{Gal}^{-/-}$ pigs in addition to the immortalized cell lines. We are aware that endothelial cells may undergo senescence by numerous passages in vitro. However, these cells have been cultured up to passage 25 without difficulties and with retained typical endothelial morphology and phenotype, including expression of receptors such as CD31, CD54, CD62E, CD62P, and CD106 (24). Furthermore, the cell lines were examined and compared at the same number of passages. The physiologic relevance of the $\text{Gal}^{-/-}$ immortalized cells used by our group and others have been questioned because they lack VCAM-1 (CD106) in their resting state (24, 42), a molecule important for adhesion of PBMC (43, 44). However, there have been technical difficulties in detecting up-regulation of VCAM-1 even on activated naive $\text{Gal}^{+/+}$ PAECs,

probably because good anti-porcine VCAM-1 Abs, to our knowledge, are not commercially available. One concern using primary cells is that they cannot be cultured indefinitely, and may alter properties over short periods of time. The primary cells used in this study are used from passage 15 to 20. This may be somewhat longer than generally accepted, but even at these passage numbers, the primary PAECs exhibited virtually the same results as the immortalized cells lines (Fig. 6) and displayed an EC-typical phenotype including the expression of adhesion receptors important for human leukocyte adhesion (24). The observations that the immortalized cells in this study showed the same results as the primary PAECs, lends support to the validity of previous results obtained using these cell lines as a reliable model for *in vitro* studies on the role of Gal in pig-to-human xenotransplantation (16, 24, 45), where the immortalized Gal^{-/-} were used. The fact that immortalized and primary Gal^{+/+} PAEC induced comparable cytokine production patterns in human cells, except for only three cytokines (IFN- γ , TNF- α , and VEGF) exclusively detected when using immortalized cells, support the assumption that the primary and immortalized cells differ only slightly phenotypically, and that these cytokines might not be physiologically relevant in a clinically setting. Compared with the serum-based model we previously used (16), the present whole blood model is physiologically more relevant and allows us to explore cellular mechanisms in the mutual interaction between PAECs and blood cells.

Inflammatory cytokines have been associated with AHXR of xenografts (20), and Al-Mohanna et al. (46) demonstrated that neutrophils are one of the major sources of cytokines, even in the absence of xenoantibodies and complement, and that this cytokine release was independent of Gal structures. Our results definitely indicate that Gal has a dominant role in the induction of cytokine production in the presence of xenoantibodies, and that this cytokine response is significantly higher than that mediated by direct cellular interactions, as studied by Al-Mohanna et al. (46). The whole blood model, however, does not discriminate between the contributions of the different cells present, but rather reflects the total amount of cytokines produced by the cells present. A major advantage with the present model is the opportunity to selectively detect and quantify both human and porcine cytokines and chemokines in the same sample. A broad panel of human cytokines was found not to cross-react with pig, whereas the number of porcine-specific cytokines was more limited, mainly due to fewer available reagents.

IL-8 serves as a potent chemoattractant for granulocytes to the site of inflammation and is associated with EC activation (47, 48), and the level of porcine IL-8 correlates with E-selectin expression in the current model (own unpublished data). In the present study, substantial differences were seen between Gal^{+/+} and Gal^{-/-} PAECs both with regard to human and porcine IL-8 production, consistent with complement activation induced by anti-Gal Abs leading to C5b-9 deposition on the PAECs and C5a release to the fluid phase, both effects known to mediate IL-8 release. IL-6 is a prototypic proinflammatory cytokine which has been associated with rejection of cardiac allo- (49) and xenograft (20, 50). We found that the Gal epitope was also critically important for the release of both human and porcine IL-6, with low levels induced by Gal^{-/-} cells.

Substantially higher levels of human IFN- γ were found in supernatants of Gal^{+/+} PAECs incubated with human whole blood compared with Gal^{-/-} PAECs. Whether IFN- γ promotes or protects against xenograft rejection is unclear. A study by Wang et al. (51) found that IFN- γ together with IL-12 was protective against AHXR, whereas other studies have demonstrated that high levels

of IFN- γ are associated with rejection (52–55). The precise role of IFN- γ in xenotransplantation remains to be elucidated.

The role of IL-1 in xenograft rejection was demonstrated by the finding that treatment with an IL-1ra reduced the expression of a number of inflammatory genes and increased graft survival in rats receiving guinea pig hearts (56). In our study, we recorded no up-regulation of human IL-1, but interestingly IL-1ra was detected and the concentration was equally or slightly stronger induced by Gal^{-/-} than Gal^{+/+} PAECs, suggesting a possible endogenous protection against Gal^{-/-} graft failure.

In accordance with our observations, Solomon et al. (57) found up-regulation of the β chemokines such RANTES, MIP-1 α , MIP-1 β , and eotaxin in a pig-to-murine islet transplantation model. In our study, their human counterparts showed significantly higher levels in supernatants from Gal^{+/+} PAECs compared with those from Gal^{-/-} PAECs, indicating that Gal is important for chemokine-induced inflammation. In contrast, up-regulation of human MCP-1 was not mediated by the Gal epitope in our study, whereas increased MCP-1 levels have been associated with rejection in other xenotransplantation models (20, 57). Up-regulation of the T cell-recruiting chemokine IP-10 has been associated with xenograft rejection in several models (19, 57, 58), although a recent study showed no change in IP-10 (59). Notably, we found that human IP-10, in contrast to all other chemokines, was produced to a greater extent when blood was incubated with Gal^{-/-} PAECs than when Gal^{+/+} PAECs were used, indicating that other factors than Gal was responsible for the induction of IP-10. Importantly, IP-10 was the only chemokine which was not reduced by complement inhibition using Gal^{+/+} cells (see Fig. 5B). This implies that IP-10 is induced by a different, complement-independent, mechanism, which is enhanced on cells lacking the Gal epitope. Further investigations are needed to reveal the role of IP-10 in xenotransplantation.

The role of growth factors in xenotransplantation is unknown, while in allotransplantation both VEGF and PDGF are associated with graft rejection (60, 61). We could not document any difference in VEGF or PDGF concentrations when eliminating the Gal epitope on PAECs stimulated with human whole blood.

The mechanism of complement-mediated induction of cytokines and chemokines was revealed by inhibition of C3 activation, using the C3 inhibitor compstatin, and by blocking the C5aR using a C5aR antagonist. Consistent with previous data from the serum model, Gal^{+/+} PAEC activation was independent of C5a and mediated by the C5b-9 complex (16). In contrast, activation of human blood cells and production of human cytokines and chemokines was completely dependent on C5a, consistent with the role of C5a in induction of inflammation induced by leukocytes in whole human blood. The minor up-regulation of E-selectin on Gal^{-/-} PAECs stimulated with human whole blood seemed to be independent of complement; however, we do not exclude a possible inhibition of complement if the sensitivity of the assay was higher. Similarly, we cannot exclude a certain effect of anti-non-Gal Abs in the up-regulation of cytokines in a Gal^{-/-} system, which might have passed undetected in our experiments due to the sensitivity of the assay. In any case, our data indicate that naturally occurring anti-non-Gal Abs play a minor role compared with anti-Gal Abs for this response.

In conclusion, we demonstrate a crucial role for Gal in xenoantibody-dependent cellular production of both porcine and human IL-6 and IL-8 in an *in vitro* model of xenograft rejection. Furthermore, human cytokines and chemokines, including IFN- γ , MIP-1 α , MIP-1 β , and eotaxin, were highly dependent on Gal. The present model and current findings contribute to further understanding of the inflammatory mechanisms involved in rejection of

cells and organs from Gal^{+/+} vs Gal^{-/-} pigs and can be used for subsequent studies to evaluate the compatibility of the next generation of gene-targeted pigs intended for human transplantation.

Disclosures

The authors have no financial conflict of interest.

References

- Galili, U., S. B. Shohet, E. Kobrin, C. L. Stults, and B. A. Macher. 1988. Man, apes, and Old World monkeys differ from other mammals in the expression of α -galactosyl epitopes on nucleated cells. *J. Biol. Chem.* 263: 17755–17762.
- Oriol, R., Y. Ye, E. Koren, and D. K. Cooper. 1993. Carbohydrate antigens of pig tissues reacting with human natural antibodies as potential targets for hyperacute vascular rejection in pig-to-man organ xenotransplantation. *Transplantation* 56: 1433–1442.
- Cooper, D. K., A. H. Good, E. Koren, R. Oriol, A. J. Malcolm, R. M. Ippolito, F. A. Neethling, Y. Ye, E. Romano, and N. Zuhdi. 1993. Identification of α -galactosyl and other carbohydrate epitopes that are bound by human anti-pig antibodies: relevance to discordant xenografting in man. *Transpl. Immunol.* 1: 198–205.
- Dalmasso, A. P., G. M. Vercellotti, R. J. Fischel, R. M. Bolman, F. H. Bach, and J. L. Platt. 1992. Mechanism of complement activation in the hyperacute rejection of porcine organs transplanted into primate recipients. *Am. J. Pathol.* 140: 1157–1166.
- McCurry, K. R., D. L. Kooyman, C. G. Alvarado, A. H. Cotterell, M. J. Martin, J. S. Logan, and J. L. Platt. 1995. Human complement regulatory proteins protect swine-to-primate cardiac xenografts from humoral injury. *Nat. Med.* 1: 423–427.
- Cozzi, E., and D. J. White. 1995. The generation of transgenic pigs as potential organ donors for humans. *Nat. Med.* 1: 964–966.
- Lai, L., D. Kolber-Simonds, K. W. Park, H. T. Cheong, J. L. Greenstein, G. S. Im, M. Samuel, A. Bonk, A. Rieke, B. N. Day, et al. 2002. Production of α -1,3-galactosyltransferase knockout pigs by nuclear transfer cloning. *Science* 295: 1089–1092.
- Phelps, C. J., C. Koike, T. D. Vaught, J. Boone, K. D. Wells, S. H. Chen, S. Ball, S. M. Specht, I. A. Polejaeva, J. A. Monahan, et al. 2003. Production of α 1,3-galactosyltransferase-deficient pigs. *Science* 299: 411–414.
- Platt, J. L., S. S. Lin, and C. G. McGregor. 1998. Acute vascular rejection. *Xenotransplantation* 5: 169–175.
- Rieben, R., and J. D. Seebach. 2005. Xenograft rejection: IgG1, complement and NK cells team up to activate and destroy the endothelium. *Trends Immunol.* 26: 2–5.
- Kuwaki, K., Y. L. Tseng, F. J. Dor, A. Shimizu, S. L. Houser, T. M. Sanderson, C. J. Lancos, D. D. Prabharasuth, J. Cheng, K. Moran, et al. 2005. Heart transplantation in baboons using α 1,3-galactosyltransferase gene-knockout pigs as donors: initial experience. *Nat. Med.* 11: 29–31.
- Chen, G., H. Qian, T. Starzl, H. Sun, B. Garcia, X. Wang, Y. Wise, Y. Liu, Y. Xiang, L. Copeman, et al. 2005. Acute rejection is associated with antibodies to non-Gal antigens in baboons using Gal-knockout pig kidneys. *Nat. Med.* 11: 1295–1298.
- Cooper, D. K. 1998. Xenoantigens and xenoantibodies. *Xenotransplantation* 5: 6–17.
- Chen, G., H. Sun, H. Yang, D. Kubelik, B. Garcia, Y. Luo, Y. Xiang, A. Qian, L. Copeman, W. Liu, et al. 2006. The role of anti-non-Gal antibodies in the development of acute humoral xenograft rejection of hDAF transgenic porcine kidneys in baboons receiving anti-Gal antibody neutralization therapy. *Transplantation* 81: 273–283.
- Zhong, R. 2007. Gal knockout and beyond. *Am. J. Transplant.* 7: 5–11.
- Saethre, M., B. C. Baumann, M. Fung, J. D. Seebach, and T. E. Mollnes. 2007. Characterization of natural human anti-non-Gal antibodies and their effect on activation of porcine Gal-deficient endothelial cells. *Transplantation* 84: 244–250.
- Marshall, S. E., and K. I. Welsh. 2001. The role of cytokine polymorphisms in rejection after solid organ transplantation. *Genes Immun.* 2: 297–303.
- Dallman, M. J., and G. J. Clark. 1991. Cytokines and their receptors in transplantation. *Curr. Opin. Immunol.* 3: 729–734.
- Hardstedt, M., C. P. Finnegan, N. Kirchoff, K. A. Hyland, M. Wijkstrom, M. P. Murtaugh, and B. J. Hering. 2005. Post-transplant upregulation of chemokine messenger RNA in non-human primate recipients of intraportal pig islet xenografts. *Xenotransplantation* 12: 293–302.
- Candinas, D., S. Belliveau, N. Koyamada, T. Miyatake, P. Hechenleitner, W. Mark, F. H. Bach, and W. W. Hancock. 1996. T cell independence of macrophage and natural killer cell infiltration, cytokine production, and endothelial activation during delayed xenograft rejection. *Transplantation* 62: 1920–1927.
- Larsen, C. G., A. O. Anderson, E. Appella, J. J. Oppenheim, and K. Matsushima. 1989. The neutrophil-activating protein (NAP-1) is also chemotactic for T lymphocytes. *Science* 243: 1464–1466.
- Cassatella, M. A. 1995. The production of cytokines by polymorphonuclear neutrophils. *Immunol. Today* 16: 21–26.
- Millan, M. T., C. Geczy, K. M. Stuhlmeier, D. J. Goodman, C. Ferran, and F. H. Bach. 1997. Human monocytes activate porcine endothelial cells, resulting in increased E-selectin, interleukin-8, monocyte chemoattractant protein-1, and plasminogen activator inhibitor-type-1 expression. *Transplantation* 63: 421–429.
- Baumann, B. C., M. K. Schneider, B. G. Lilienfeld, M. A. Antsiferova, D. M. Rhyner, R. J. Hawley, and J. D. Seebach. 2005. Endothelial cells derived from pigs lacking Gal α (1,3)Gal: no reduction of human leukocyte adhesion and natural killer cell cytotoxicity. *Transplantation* 79: 1067–1072.
- He, Z., C. Ehrnfelt, M. Kumagai-Braesch, K. B. Islam, and J. Holgersson. 2004. Aberrant expression of α -Gal on primary human endothelium does not confer susceptibility to NK cell cytotoxicity or increased NK cell adhesion. *Eur. J. Immunol.* 34: 1185–1195.
- Peterson, M. D., R. Jin, S. Hyduk, P. Duchesneau, M. I. Cybulsky, and T. K. Waddell. 2005. Monocyte adhesion to xenogeneic endothelium during laminar flow is dependent on α -Gal-mediated monocyte activation. *J. Immunol.* 174: 8072–8081.
- Seebach, J. D., M. K. Schneider, C. A. Comrack, A. LeGuern, S. A. Kolb, P. A. Knelle, S. Germana, H. DerSimonian, C. LeGuern, and D. H. Sachs. 2001. Immortalized bone-marrow derived pig endothelial cells. *Xenotransplantation* 8: 48–61.
- Baumann, B. C., P. Forte, R. J. Hawley, R. Rieben, M. K. Schneider, and J. D. Seebach. 2004. Lack of galactose- α -1,3-galactose expression on porcine endothelial cells prevents complement-induced lysis but not direct xenogeneic NK cytotoxicity. *J. Immunol.* 172: 6460–6467.
- Holten, E. 1979. Serotypes of *Neisseria meningitidis* isolated from patients in Norway during the first six months of 1978. *J. Clin. Microbiol.* 9: 186–188.
- Sahu, A., B. K. Kay, and J. D. Lambris. 1996. Inhibition of human complement by a C3-binding peptide isolated from a phage-displayed random peptide library. *J. Immunol.* 157: 884–891.
- Katragadda, M., P. Magotti, G. Sfyroera, and J. D. Lambris. 2006. Hydrophobic effect and hydrogen bonds account for the improved activity of a complement inhibitor, compstatin. *J. Med. Chem.* 49: 4616–4622.
- Mastellos, D., J. C. Papadimitriou, S. Franchini, P. A. Tsonis, and J. D. Lambris. 2001. A novel role of complement: mice deficient in the fifth component of complement (C5) exhibit impaired liver regeneration. *J. Immunol.* 166: 2479–2486.
- Lappegard, K. T., J. Riesenfeld, O. L. Brekke, G. Bergseth, J. D. Lambris, and T. E. Mollnes. 2005. Differential effect of heparin coating and complement inhibition on artificial surface-induced eicosanoid production. *Ann. Thorac. Surg.* 79: 917–923.
- Lai, L., D. Kolber-Simonds, K. W. Park, H. T. Cheong, J. L. Greenstein, G. S. Im, M. Samuel, A. Bonk, A. Rieke, B. N. Day, et al. 2002. Production of α -1,3-galactosyltransferase knockout pigs by nuclear transfer cloning. *Science* 295: 1089–1092.
- Phelps, C. J., C. Koike, T. D. Vaught, J. Boone, K. D. Wells, S. H. Chen, S. Ball, S. M. Specht, I. A. Polejaeva, J. A. Monahan, et al. 2003. Production of α 1,3-galactosyltransferase-deficient pigs. *Science* 299: 411–414.
- Tai, H. C., M. Ezzelarab, H. Hara, D. Ayares, and D. K. Cooper. 2007. Progress in xenotransplantation following the introduction of gene-knockout technology. *Transpl. Int.* 20: 107–117.
- Cowan, P. J. 2007. Coagulation and the xenograft endothelium. *Xenotransplantation* 14: 7–12.
- Tseng, Y. L., K. Kuwaki, F. J. Dor, A. Shimizu, S. Houser, Y. Hisashi, K. Yamada, S. C. Robson, M. Awad, H. J. Schuurman, et al. 2005. α 1,3-Galactosyltransferase gene-knockout pig heart transplantation in baboons with survival approaching 6 months. *Transplantation* 80: 1493–1500.
- Markiewski, M. M., B. Nilsson, K. N. Ekdahl, T. E. Mollnes, and J. D. Lambris. 2007. Complement and coagulation: strangers or partners in crime? *Trends Immunol.* 28: 184–192.
- Mollnes, T. E., O. L. Brekke, M. Fung, H. Fure, D. Christiansen, G. Bergseth, V. Videm, K. T. Lappegard, J. Kohl, and J. D. Lambris. 2002. Essential role of the C5a receptor in *E. coli*-induced oxidative burst and phagocytosis revealed by a novel lepirudin-based human whole blood model of inflammation. *Blood* 100: 1869–1877.
- Banz, Y., T. Cung, E. Y. Korchagina, N. V. Bovin, A. Haerberli, and R. Rieben. 2005. Endothelial cell protection and complement inhibition in xenotransplantation: a novel in vitro model using whole blood. *Xenotransplantation* 12: 434–443.
- Baumann, B. C., P. Forte, R. J. Hawley, R. Rieben, M. K. Schneider, and J. D. Seebach. 2004. Lack of galactose- α -1,3-galactose expression on porcine endothelial cells prevents complement-induced lysis but not direct xenogeneic NK cytotoxicity. *J. Immunol.* 172: 6460–6467.
- Robinson, L. A., L. Tu, D. A. Steeber, O. Preis, J. L. Platt, and T. F. Tedder. 1998. The role of adhesion molecules in human leukocyte attachment to porcine vascular endothelium: implications for xenotransplantation. *J. Immunol.* 161: 6931–6938.
- Schneider, M. K., M. Strasser, U. O. Gilli, M. Kocher, R. Moser, and J. D. Seebach. 2002. Rolling adhesion of human NK cells to porcine endothelial cells mainly relies on CD49d-CD106 interactions. *Transplantation* 73: 789–796.
- Baumann, B. C., P. Forte, R. J. Hawley, R. Rieben, M. K. Schneider, and J. D. Seebach. 2004. Lack of galactose- α -1,3-galactose expression on porcine endothelial cells prevents complement-induced lysis but not direct xenogeneic NK cytotoxicity. *J. Immunol.* 172: 6460–6467.
- Al-Mohanna, F., S. Saleh, R. S. Parhar, K. Khabar, and K. Collison. 2005. Human neutrophil gene expression profiling following xenogeneic encounter with porcine aortic endothelial cells: the occult role of neutrophils in xenograft rejection revealed. *J. Leukocyte Biol.* 78: 51–61.
- Goodman, D. J., A. M. Von, A. Willson, M. T. Millan, and F. H. Bach. 1996. Direct activation of porcine endothelial cells by human natural killer cells. *Transplantation* 61: 763–771.
- Gilli, U. O., M. K. Schneider, P. Loetscher, and J. D. Seebach. 2005. Human polymorphonuclear neutrophils are recruited by porcine chemokines acting on

- CXC chemokine receptor 2, and platelet-activating factor. *Transplantation* 79: 1324–1331.
49. Deng, M. C., G. Plenz, C. Labarrere, C. Marboe, H. A. Baba, M. Erre, and S. Itescu. 2002. The role of IL6 cytokines in acute cardiac allograft rejection. *Transpl. Immunol.* 9: 115–120.
50. Matsumiya, G., S. R. Gundry, S. Nehlsen-Cannarella, O. R. Fagoaga, T. Morimoto, S. Arai, N. Fukushima, C. W. Zuppan, and L. L. Bailey. 1997. Serum interleukin-6 level after cardiac xenotransplantation in primates. *Transplant. Proc.* 29: 916–919.
51. Wang, H., M. E. DeVries, S. Deng, M. H. Khandaker, J. G. Pickering, L. H. Chow, B. Garcia, D. J. Kelvin, and R. Zhong. 2000. The axis of interleukin 12 and γ interferon regulates acute vascular xenogeneic rejection. *Nat. Med.* 6: 549–555.
52. Tanaka, K., S. Yamagami, and J. W. Streilein. 2005. Evidence that T-helper type 2 cell-derived cytokines and eosinophils contribute to acute rejection of orthotopic corneal xenografts in mice. *Transplantation* 79: 1317–1323.
53. Holan, V., J. Pindjakova, A. Zajicova, M. Krulova, B. Zelezna, P. Matousek, and P. Svoboda. 2005. The activity of inducible nitric oxide synthase in rejected skin xenografts is selectively inhibited by a factor produced by grafted cells. *Xenotransplantation* 12: 227–234.
54. Zhang, X. G., Y. Lu, B. Wang, H. Li, L. Yu, C. Liu, Z. Wu, and X. M. Liu. 2007. Cytokine production during the inhibition of acute vascular rejection in a concordant hamster-to-rat cardiac xenotransplantation model. *Chin. Med. J.* 120: 145–149.
55. Lorant, T., H. Krook, J. Wilton, M. Olausson, G. Tufveson, O. Korsgren, and C. Johnsson. 2003. Intragraft cytokine mRNA expression in rejecting and non-rejecting vascularized xenografts. *Xenotransplantation* 10: 311–324.
56. Saadi, S., T. Takahashi, R. A. Holzknacht, and J. L. Platt. 2004. Pathways to acute humoral rejection. *Am. J. Pathol.* 164: 1073–1080.
57. Solomon, M. F., W. A. Kuziel, D. A. Mann, and C. J. Simeonovic. 2003. The role of chemokines and their receptors in the rejection of pig islet tissue xenografts. *Xenotransplantation* 10: 164–177.
58. Lee, E. M., J. O. Park, D. Kim, J. Y. Kim, K. H. Oh, C. G. Park, B. H. Oh, S. Kim, and C. Ahn. 2006. Early up-regulation of CXC-chemokine expression is associated with strong cellular immune responses to murine skin xenografts. *Xenotransplantation* 13: 328–336.
59. Kim, J. Y., D. Kim, E. M. Lee, I. Choi, C. G. Park, K. S. Kim, J. Ha, S. J. Kim, J. Yang, Y. S. Kim, et al. 2003. Inducible nitric oxide synthase inhibitors prolonged the survival of skin xenografts through selective down-regulation of pro-inflammatory cytokine and CC-chemokine expressions. *Transpl. Immunol.* 12: 63–72.
60. Nykanen, A. I., R. Krebs, J. M. Tikkanen, O. Raisky, R. Sihvola, J. Wood, P. K. Koskinen, and K. B. Lemstrom. 2005. Combined vascular endothelial growth factor and platelet-derived growth factor inhibition in rat cardiac allografts: beneficial effects on inflammation and smooth muscle cell proliferation. *Transplantation* 79: 182–189.
61. Tambur, A. R., S. Pamboukian, M. R. Costanzo, and A. Heroux. 2006. Genetic polymorphism in platelet-derived growth factor and vascular endothelial growth factor are significantly associated with cardiac allograft vasculopathy. *J. Heart Lung Transplant.* 25: 690–698.